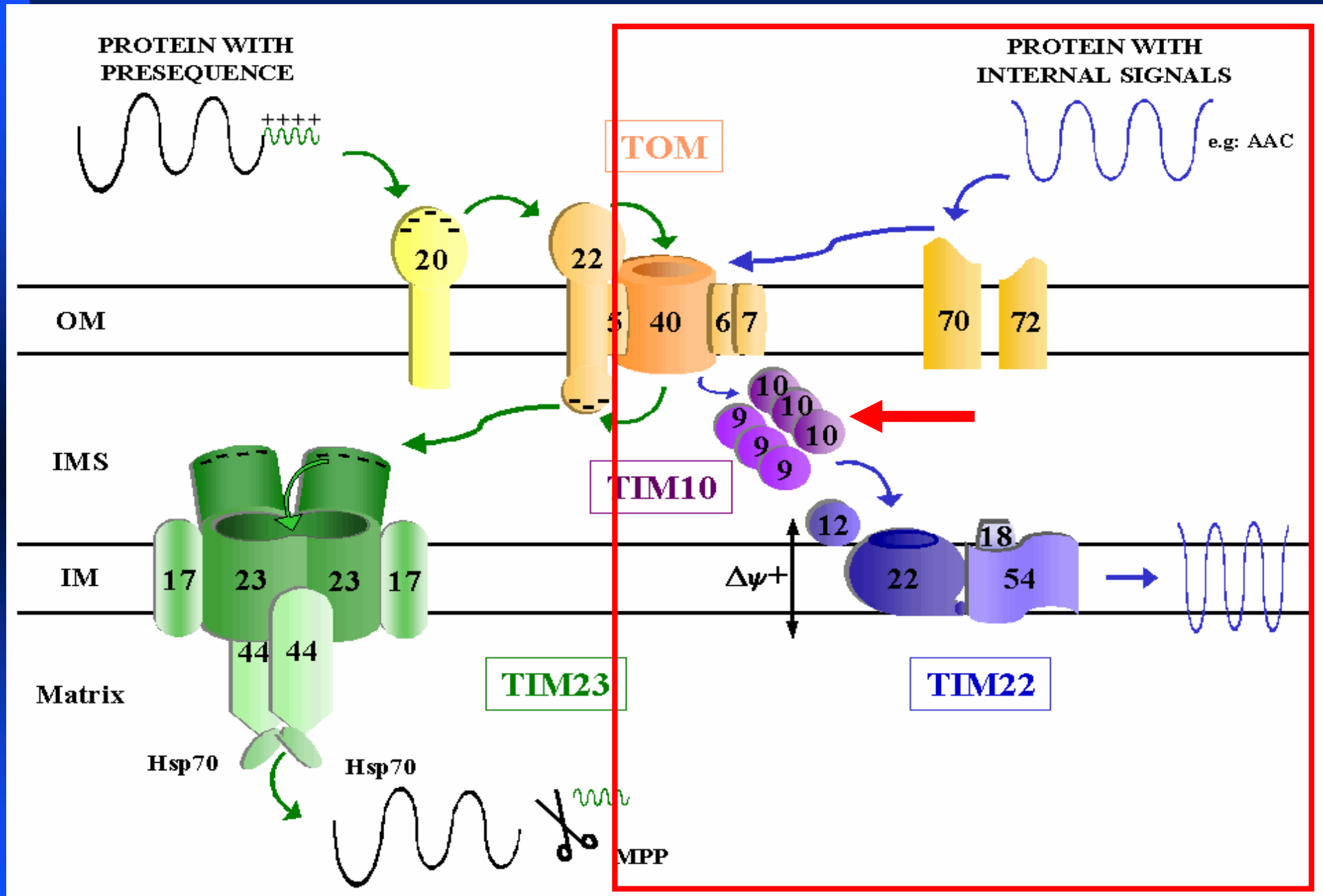


In mammalian cells mitochondrial carriers are involved in a number of disorders:

- **diseases, like several myopathies**
- **obesity**
- **programmed cell death (apoptosis)**



Distinct features of the “carrier pathway” compared to “matrix targeting”:

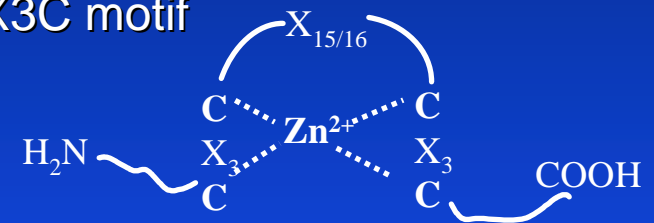
- **1. Requirement for a chaperone function in the IMS**
- **2. No ATP hydrolysis in the matrix is necessary**
- **3. No translocation motor in the matrix is required**
- **4. The electrochemical membrane potential across the IM is enough to complete insertion at the IM**

Properties of the “small Tim proteins” - 1

■ Sequence characteristics:

- ◆ They are all homologous
- ◆ They are intrinsically soluble (No transmembrane domains)
- ◆ They have a putative “zinc-binding” motif, twin CX₃C motif
- ◆ Found in all eukaryotic cells
- ◆ Human homologue of Tim8

involved in Mohr-Tranebjaerg syndrome



■ Organisation in assemblies:

- ◆ Tim9/10 and Tim8/13 are found in 70 kDa complexes in the IMS
- ◆ Tim12 is associated with the membrane-embedded TIM22 complex (IM)

Properties of the “small Tim proteins” - 2

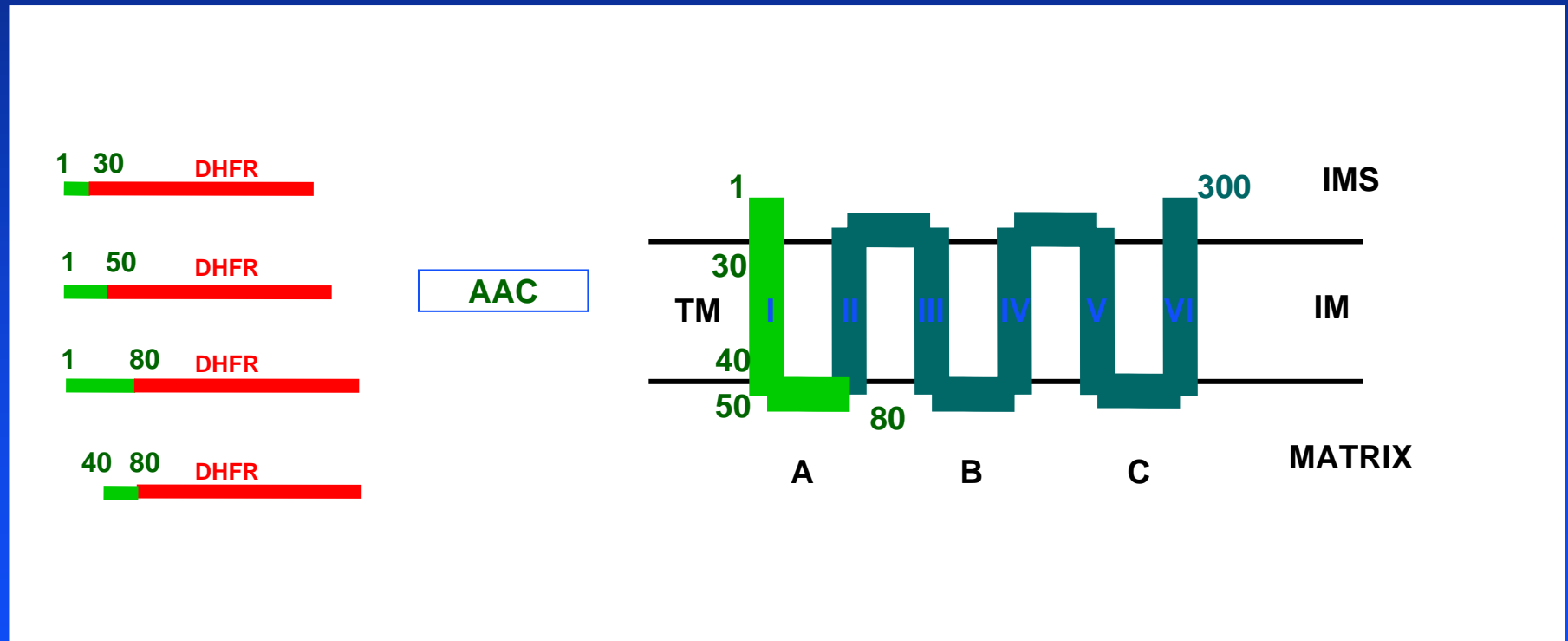
■ Function:

- ◆ Tim9/10 (essential) function as chaperones in the IMS and facilitate targeting to the IM
- ◆ Tim12 (essential) facilitates insertion in the context of the TIM22 complex
- ◆ Tim8/13 are non-essential but also seem to function as chaperones in the IMS

The AAC substrate:
targeting signals??

Which are the sorting and insertion signals of the carrier proteins?

A. Deletion Analysis

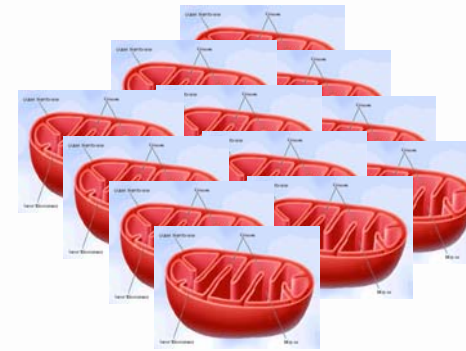
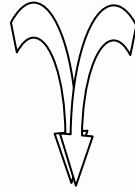


B. Site-directed mutagenesis

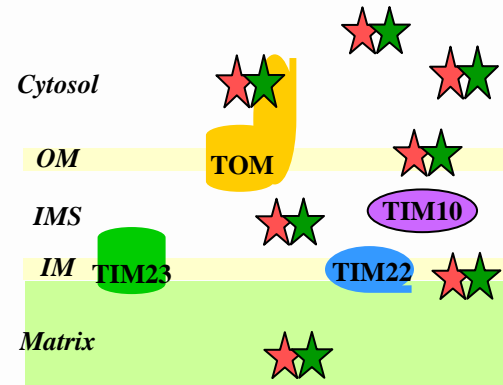


Radiolabelled protein synthesised in a rabbit reticulocyte lysate.

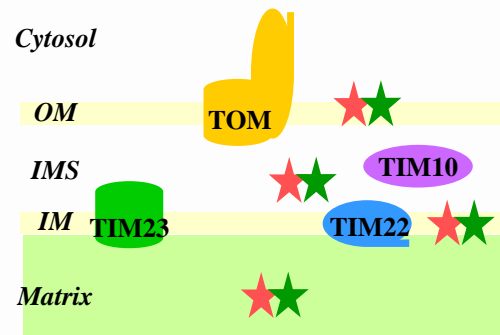
Import for
10min at
30°C



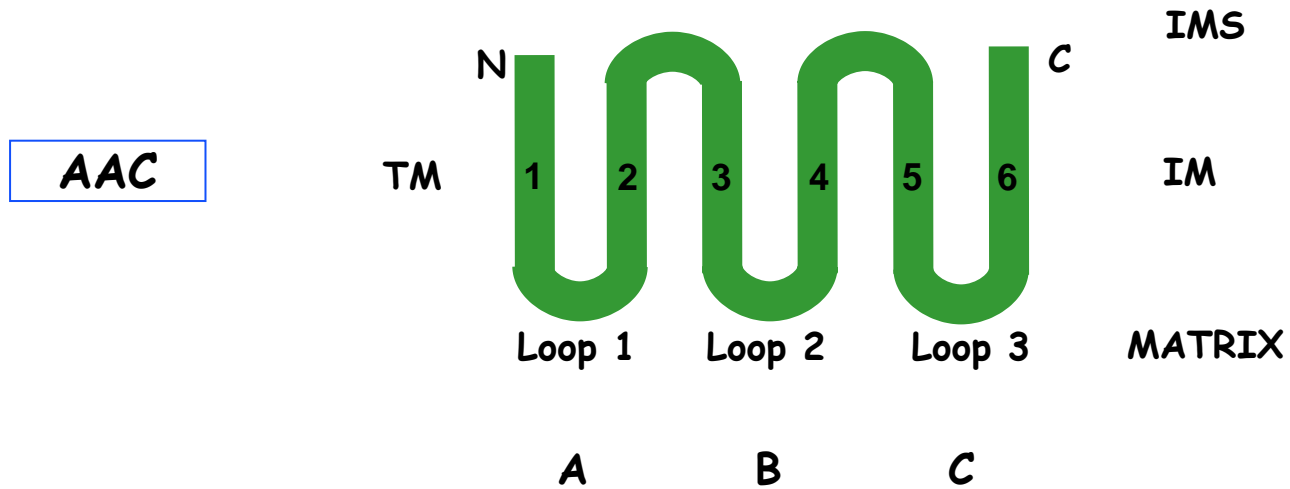
Purified
mitochondria



Proteinase
K treatment



AAC-DHFR CONSTRUCTS



AAC fragment

DHFR



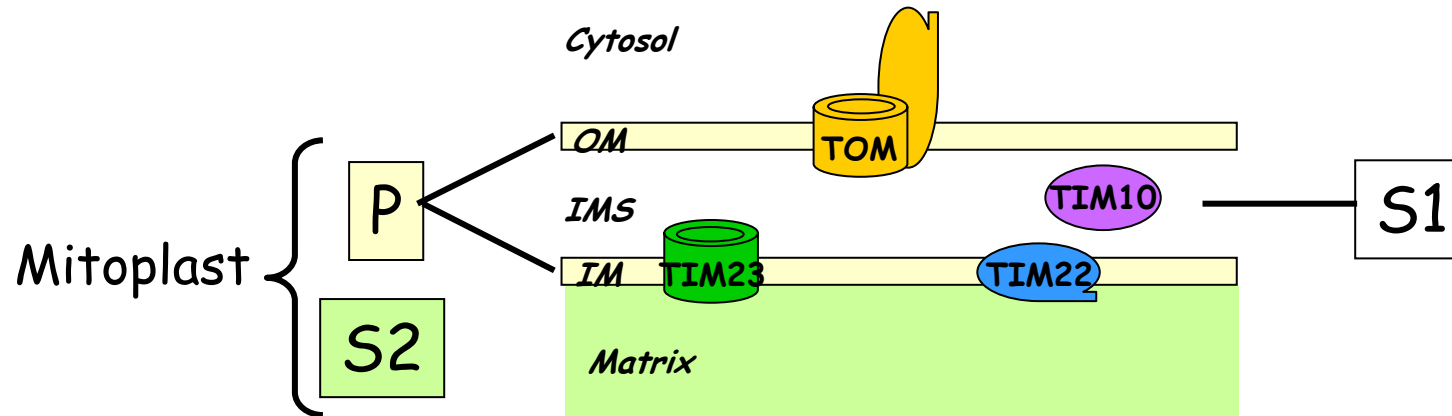
1TM

2TM

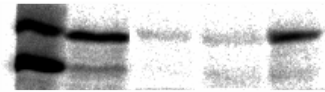
4TM

1Loop

LOCALISATION



10% tot S1 P S2



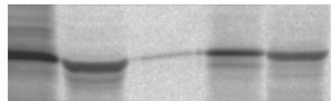
1TM



1loop



2TM

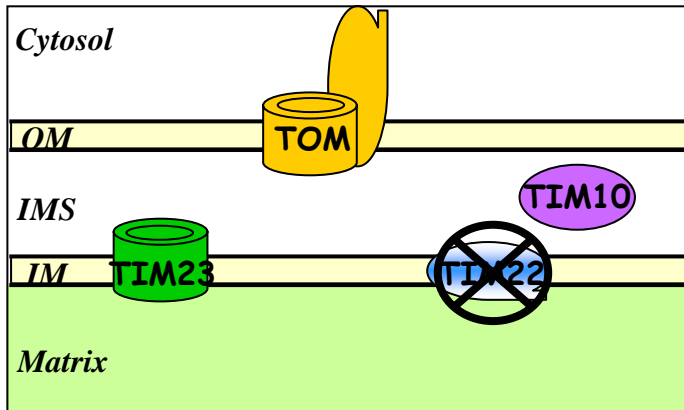


4TM

•Main localisation in IM

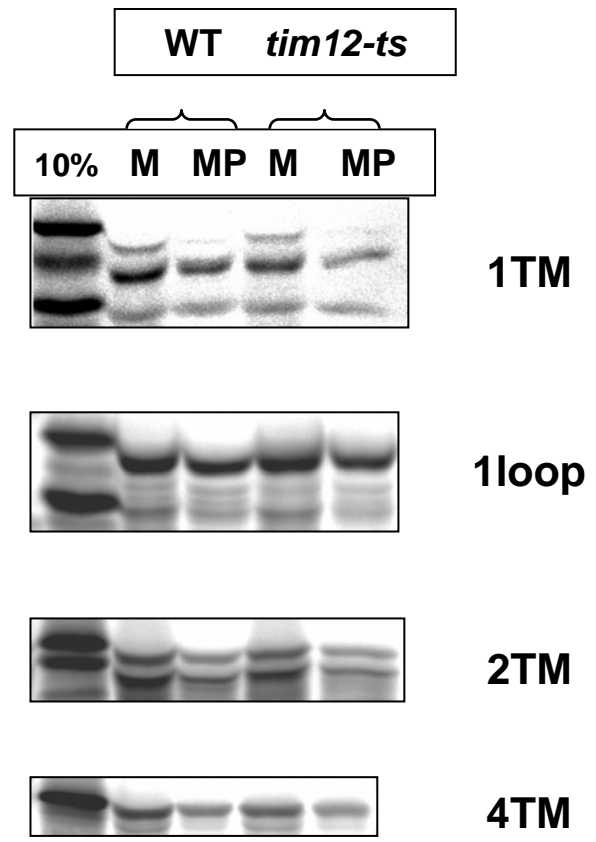
•Increase TM numbers → IM and Matrix

IS THE TIM22 COMPLEX REQUIRED FOR IMPORT?

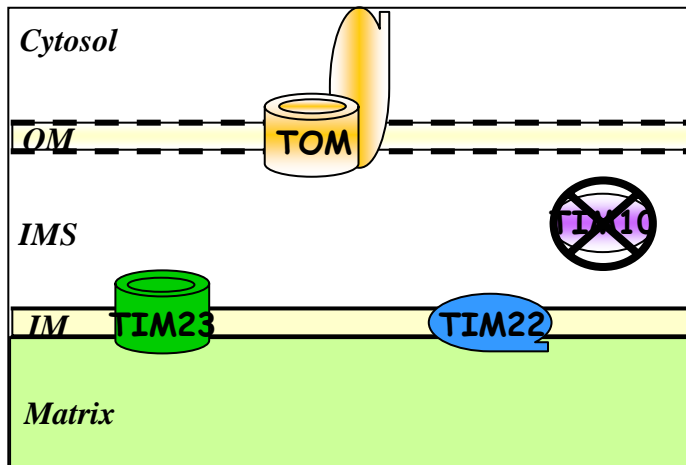


Import into *tim12-ts* mitochondria

- No Tim12
- No Tim22

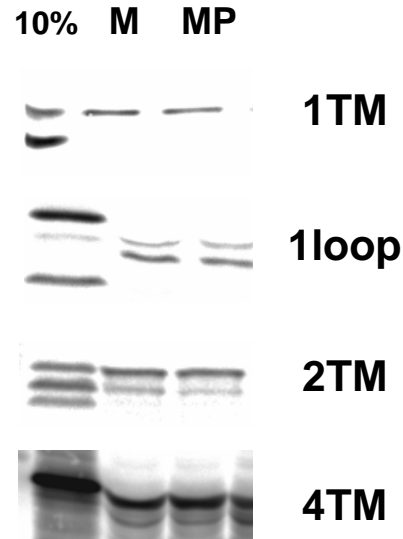


IS THE TIM10 COMPLEX REQUIRED FOR IMPORT?

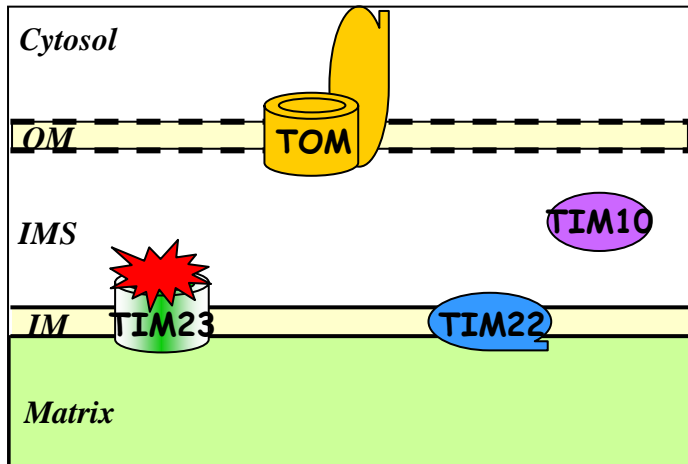


Import into Mitoplasts

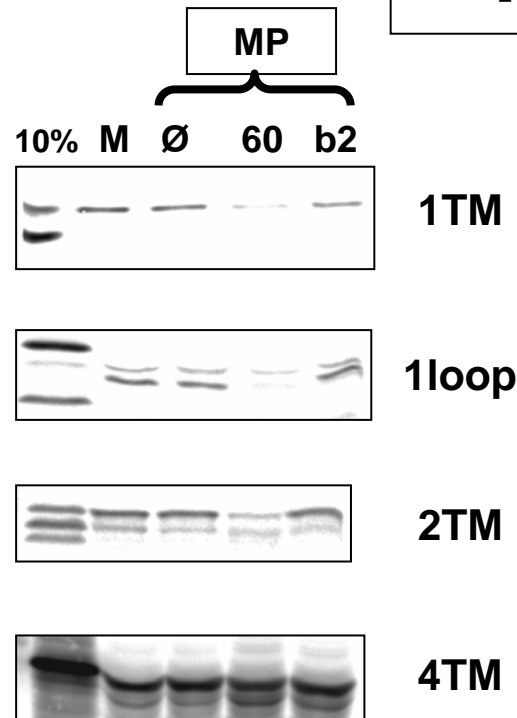
- No TIM10 complex



IS THE TIM23 COMPLEX REQUIRED FOR IMPORT?



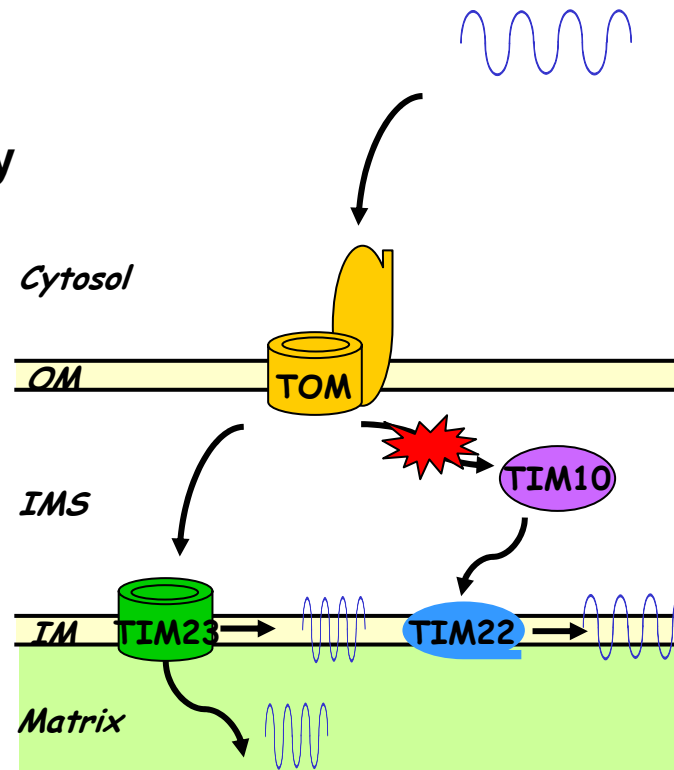
Import into mitoplasts +
peptide competition
-Hsp60= presequence peptide
- Synb2= charged peptide



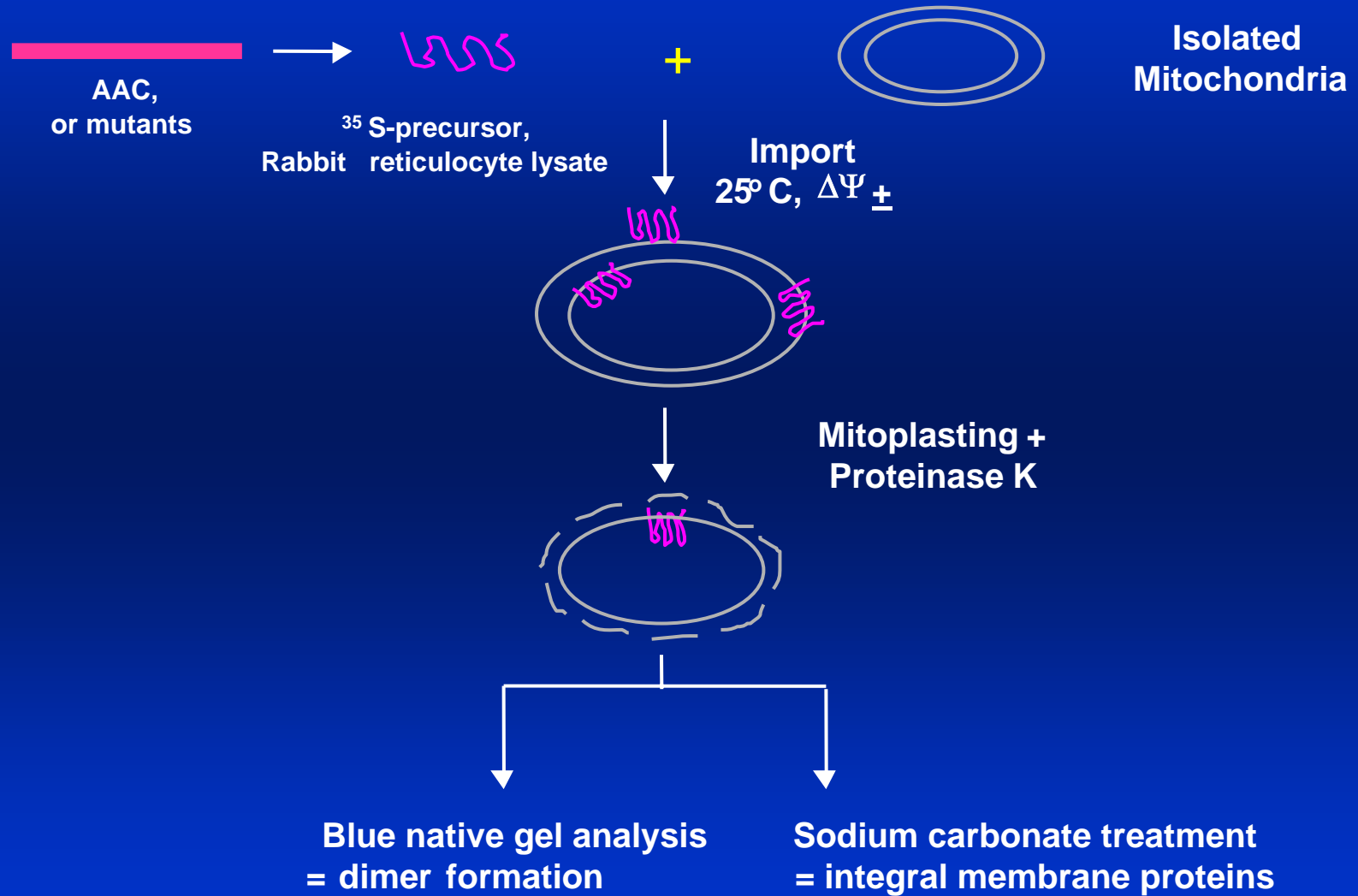
WORKING MODEL

Sufficient targeting information within each construct, BUT:

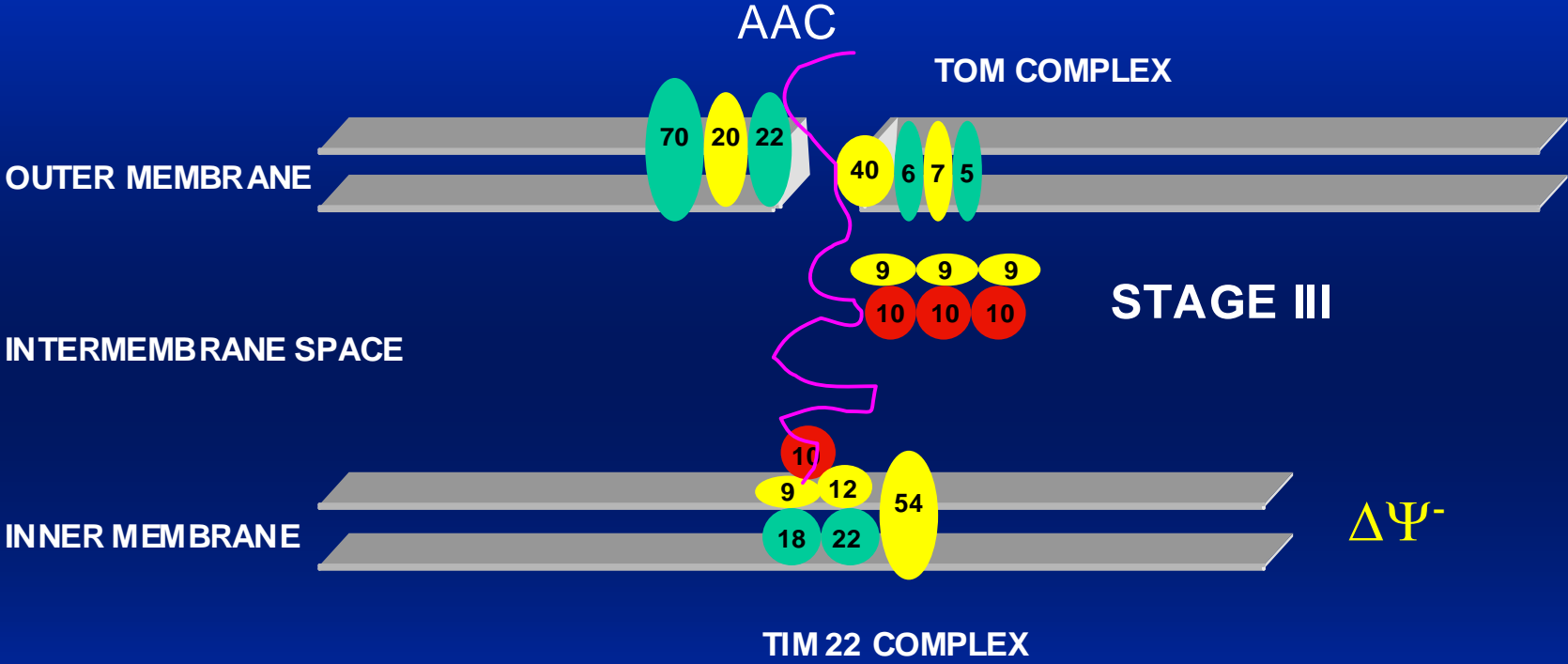
- No interaction with TIM10 complex
- Therefore no targeting to TIM22
- Translocation via TIM23 as a default pathway



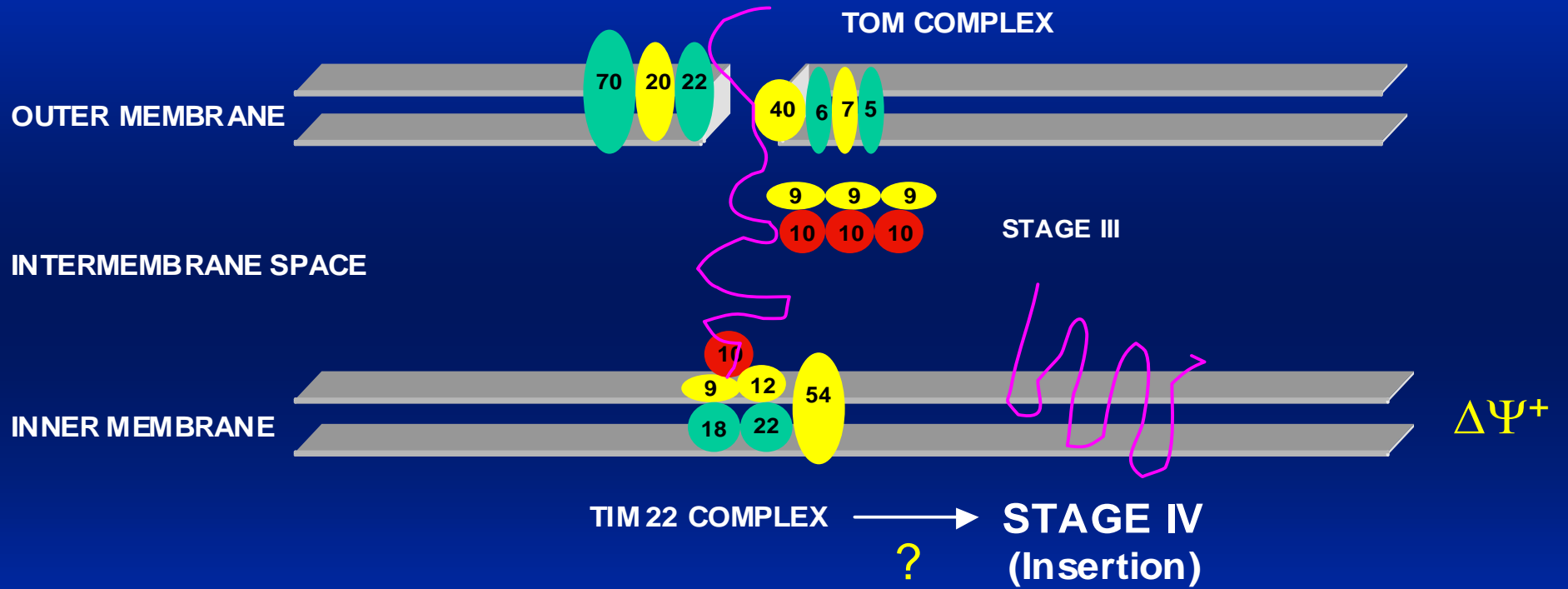
Import Scheme for AAC



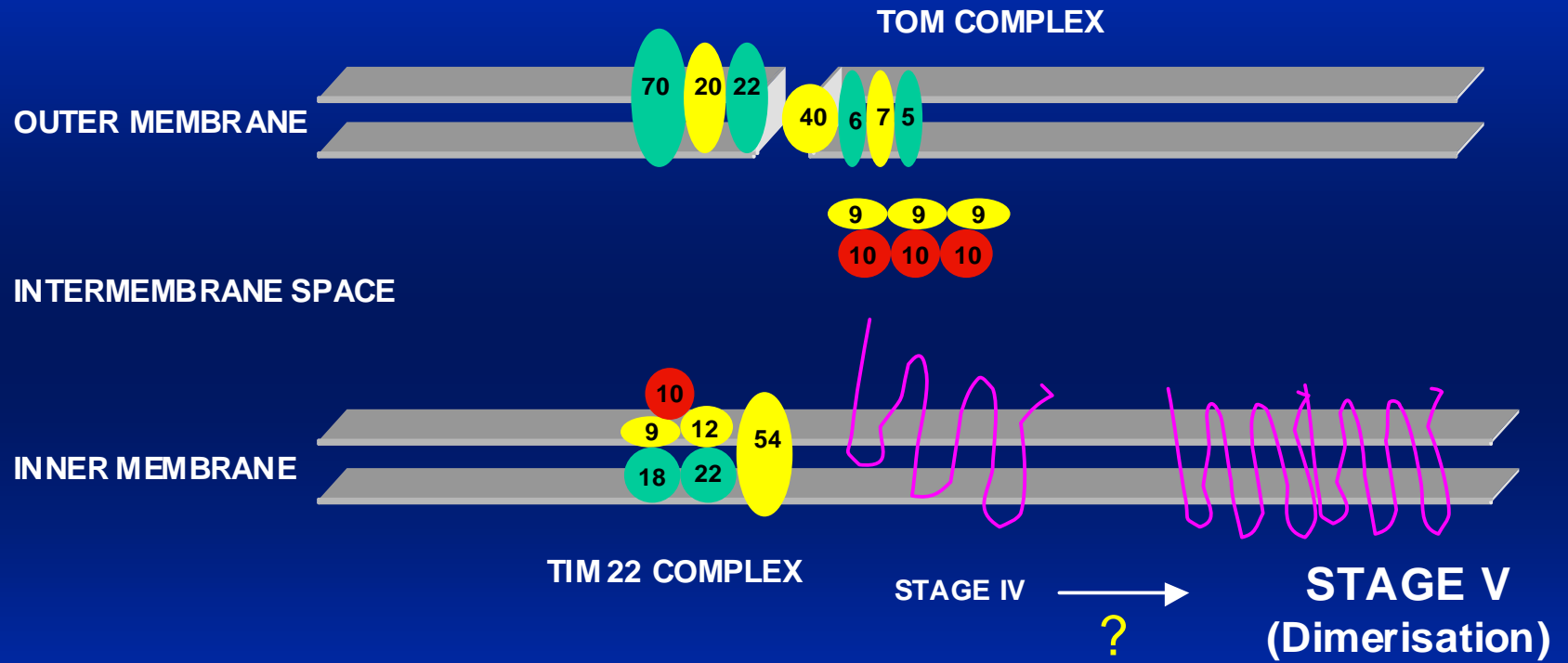
Stage III of AAC import

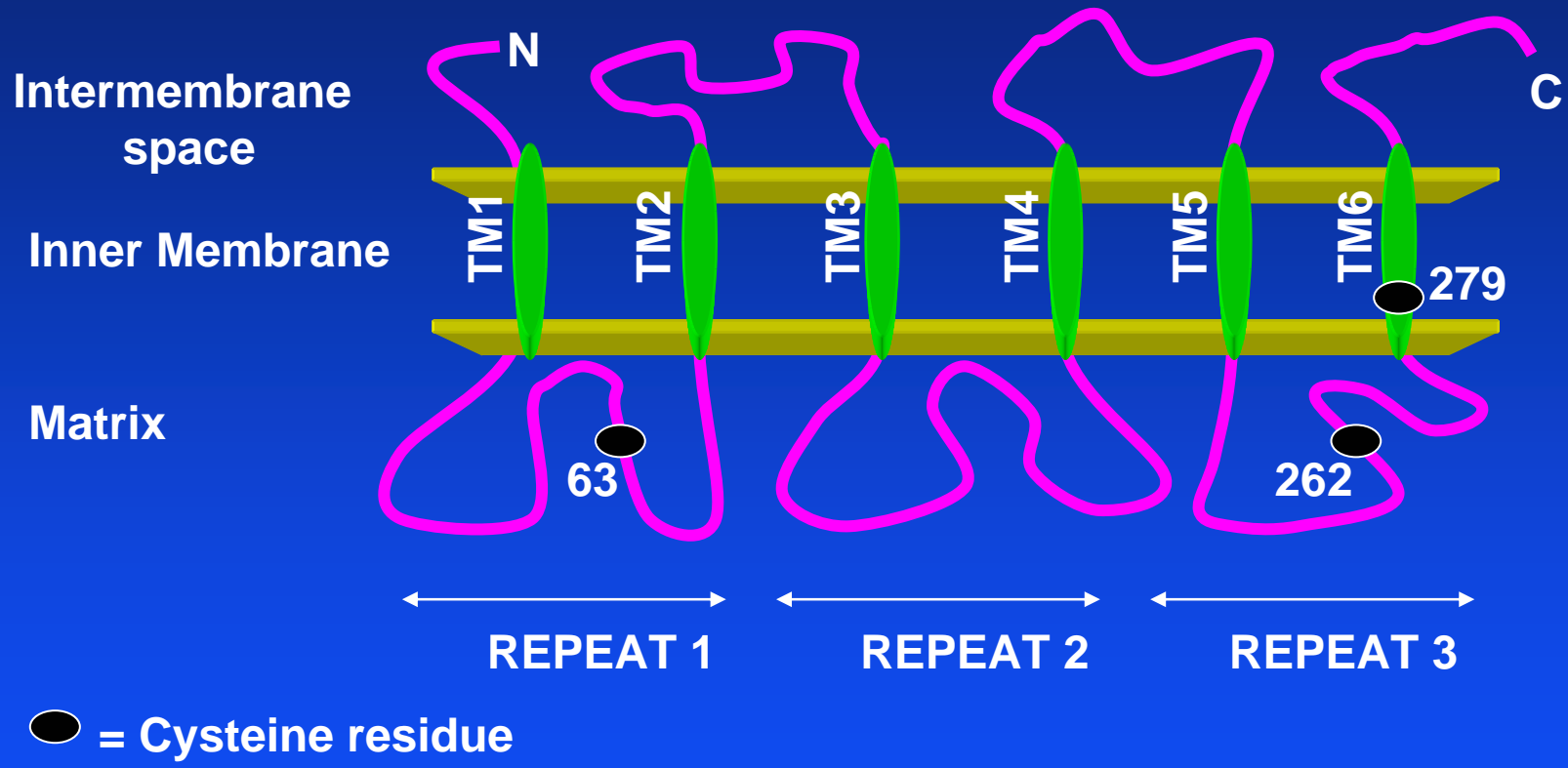


Stage IV of AAC Import

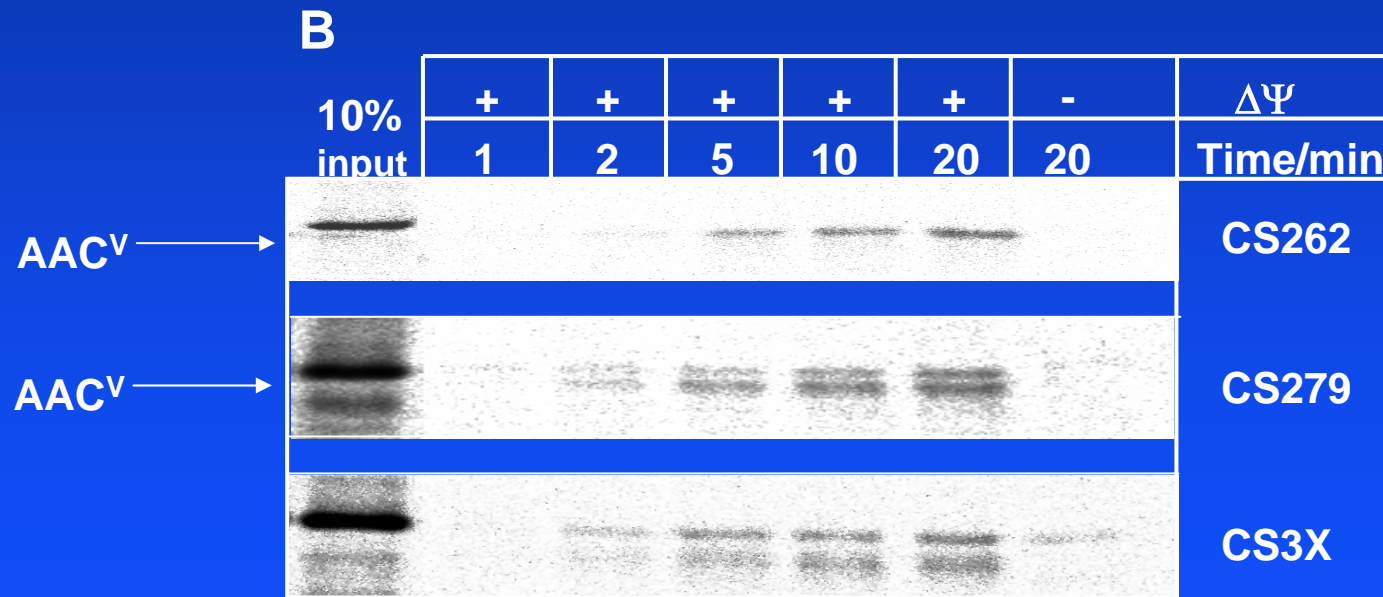
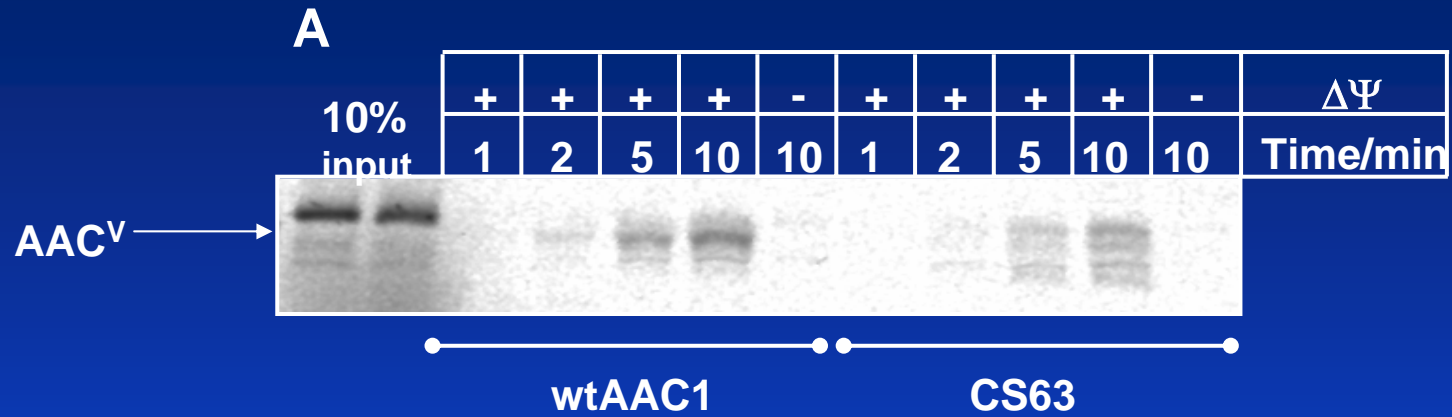


Stage V of AAC Import

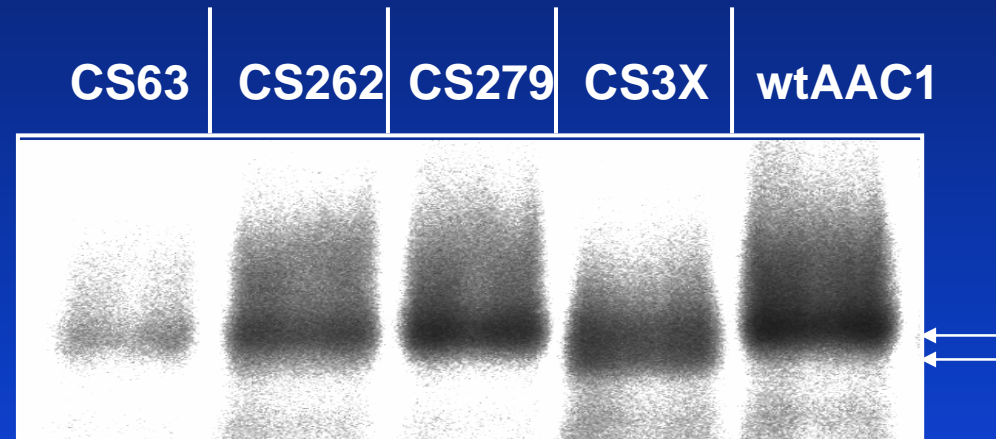




Cys mutants affecting AAC insertion in the IM



Cys mutants affecting AAC insertion in the IM



- Functionally impaired Cys mutants have a significant defect also in insertion and dimerisation of AAC1
- CS63 (first loop) and the triple Cys mutant are abrogated in their efficiency to form a dimer in the membrane

Can we reconstitute *in vitro* the TIM10 complex in a functional form?

Reconstitution of the complex from the individual subunits

1. Co-expression of Tim9 and Tim10 in *E. coli* on a single plasmid in an operon
2. Expression and purification of the individual proteins separately
3. Purification of the authentic complex from yeast mitochondria

YES!

Tim9, Tim10 necessary and sufficient for assembly of TIM10 complex

Reconstituted complex indistinguishable from mitochondrial complex

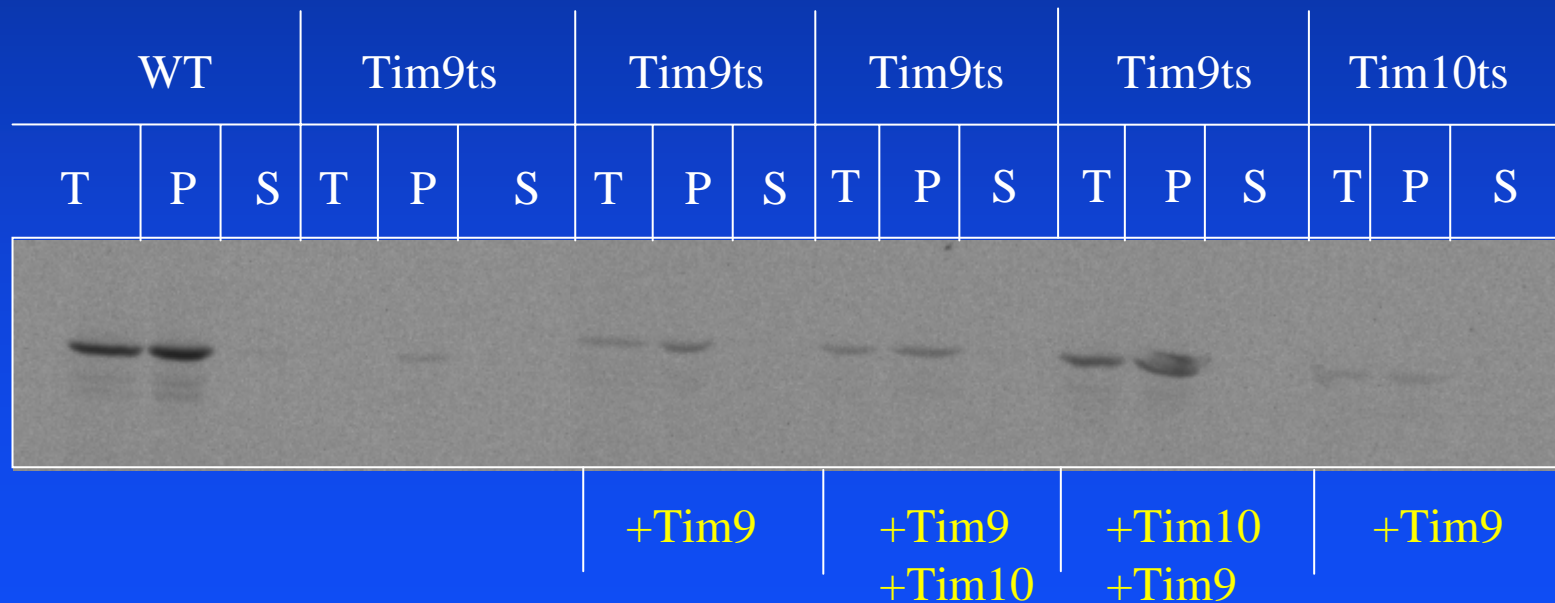
Is the reconstituted complex functional?

Functionality Assays:

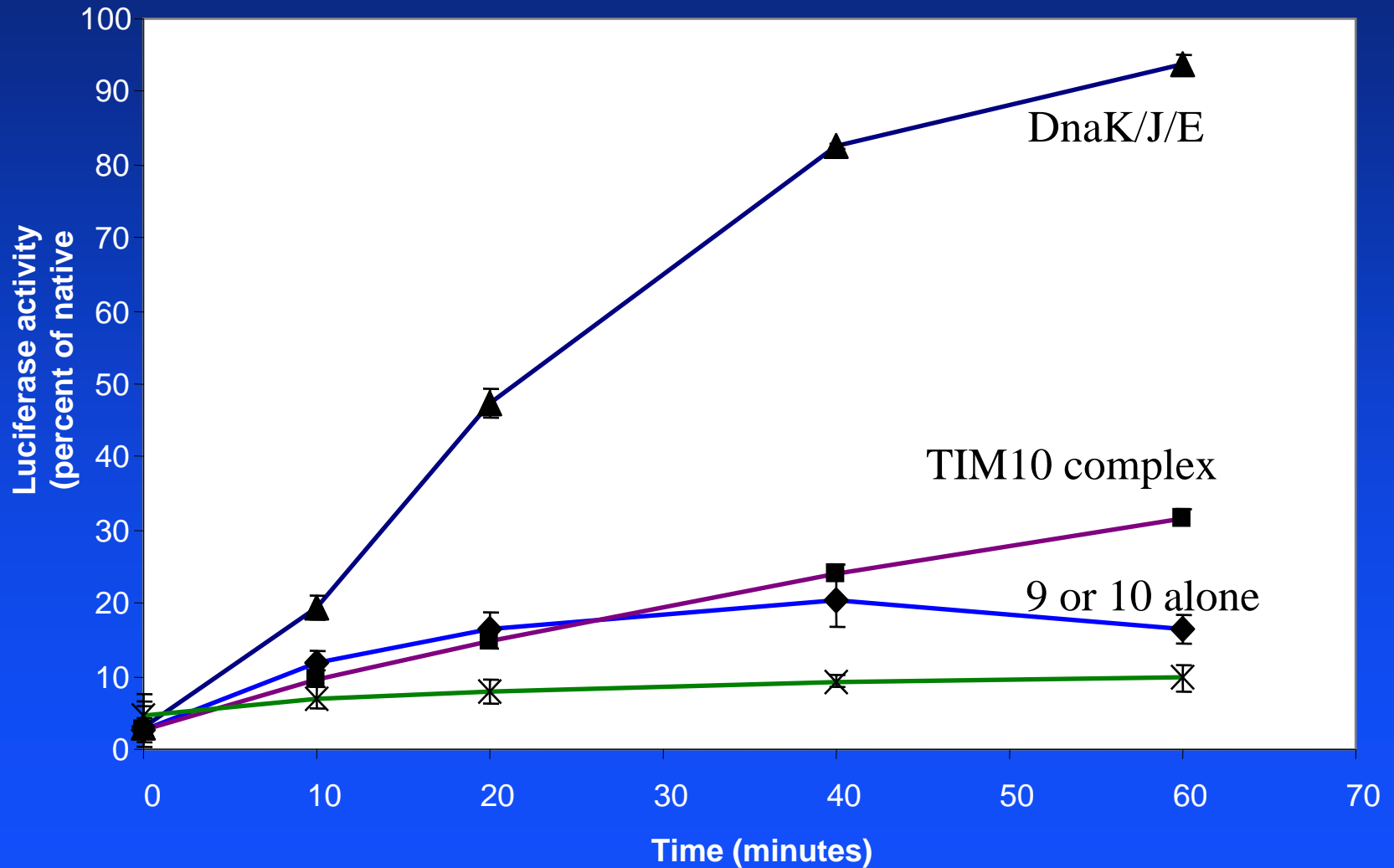
1. Restoration of AAC import into TIM10-depleted mitochondria
2. Binding in vitro to AAC
3. Chaperone activity in vitro

Reconstitution in Tim9ts Mitochondria

AAC import into Tim9ts Mitochondria
is restored after Import of Tim9 and/or Tim10

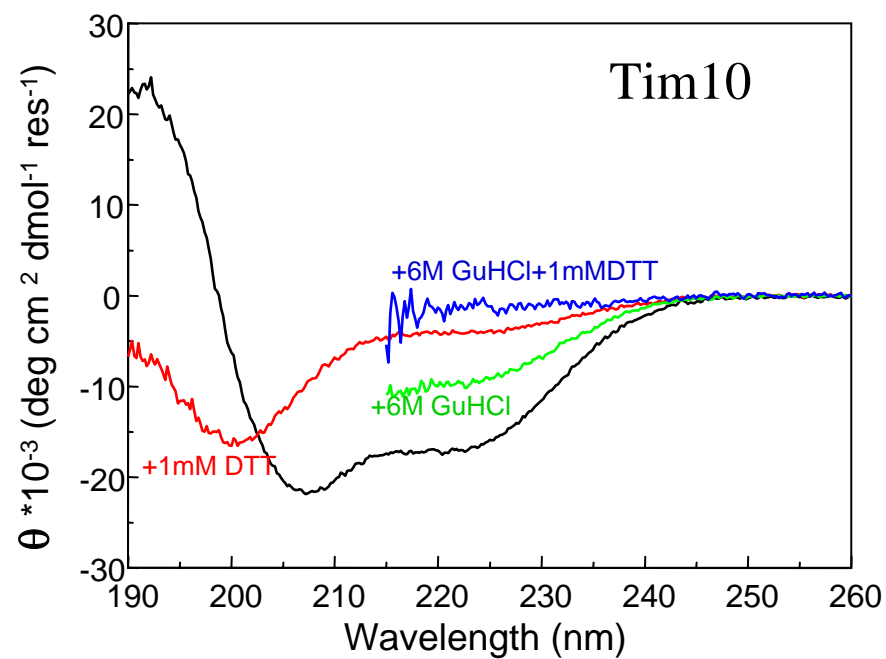
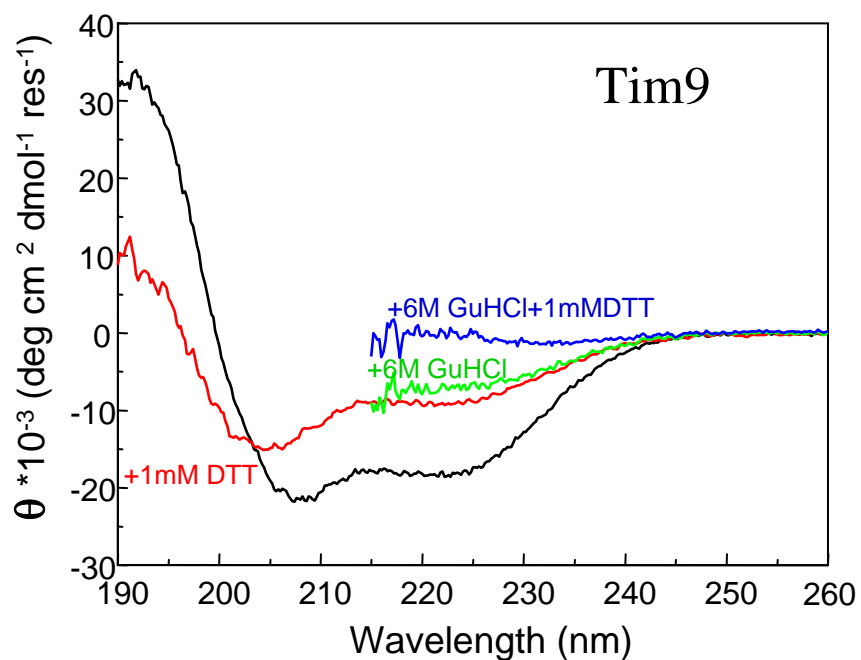


The TIM10 complex chaperones luciferase refolding



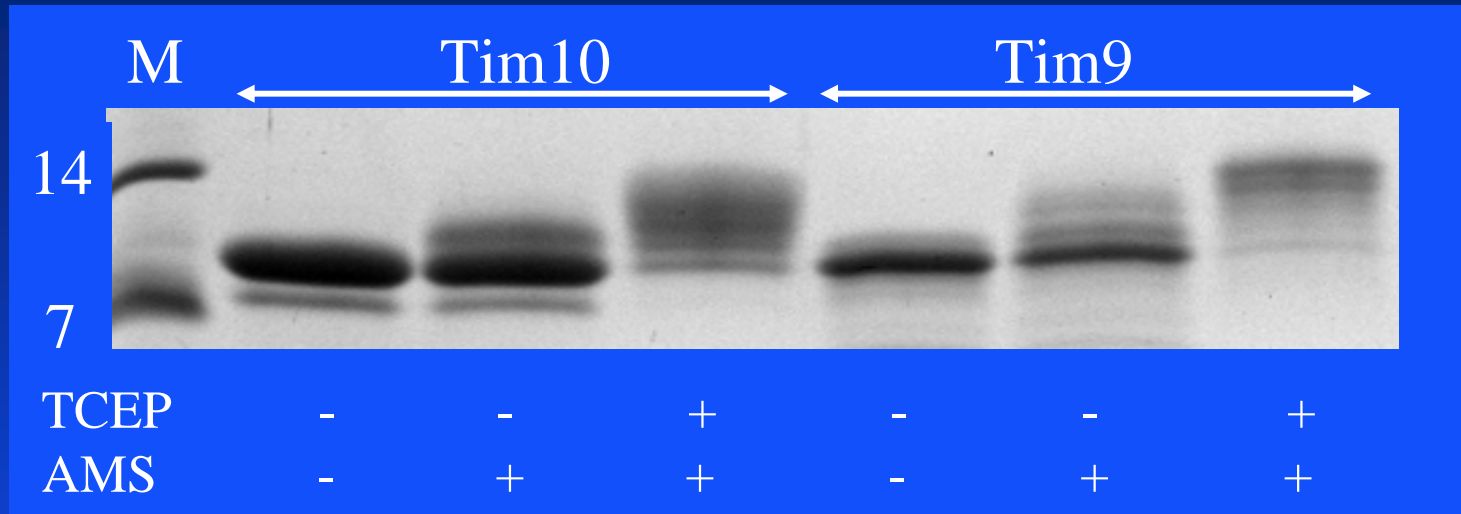
What is the structural basis for the assembly of the TIM10 complex?

CD Analysis of individual Tim9 and Tim10



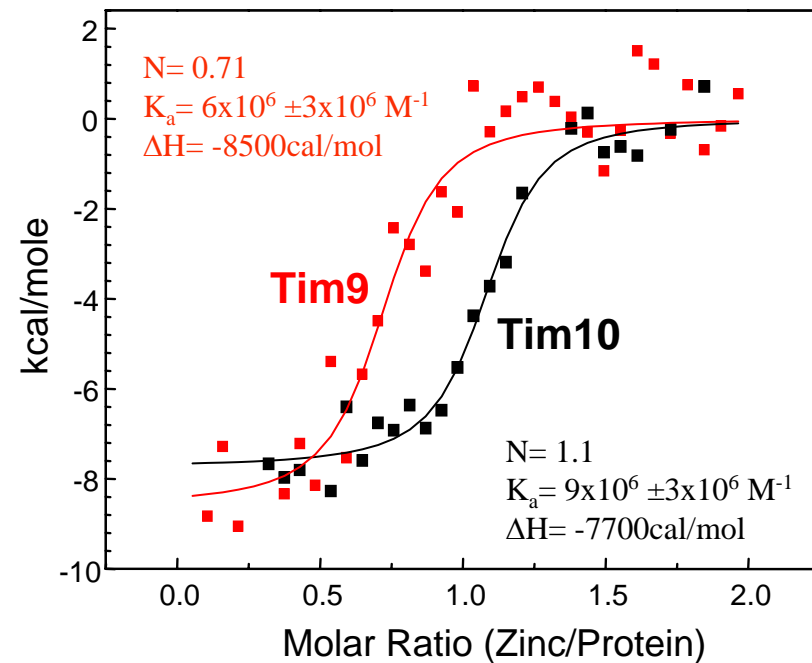
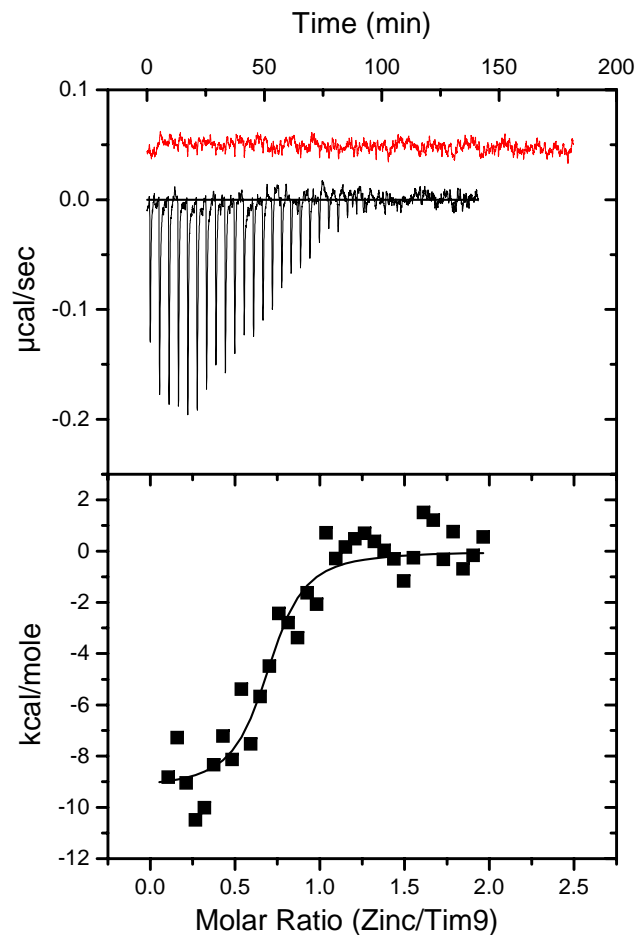
	α -helix	β -sheet	rest
oxTim10	65%	-	35%
red Tim10	8%	33%	59%
oxTim9	53%	8%	38%
redTim9	12%	39%	49%

Thiol-trapping and accessibility by AMS and DTNB

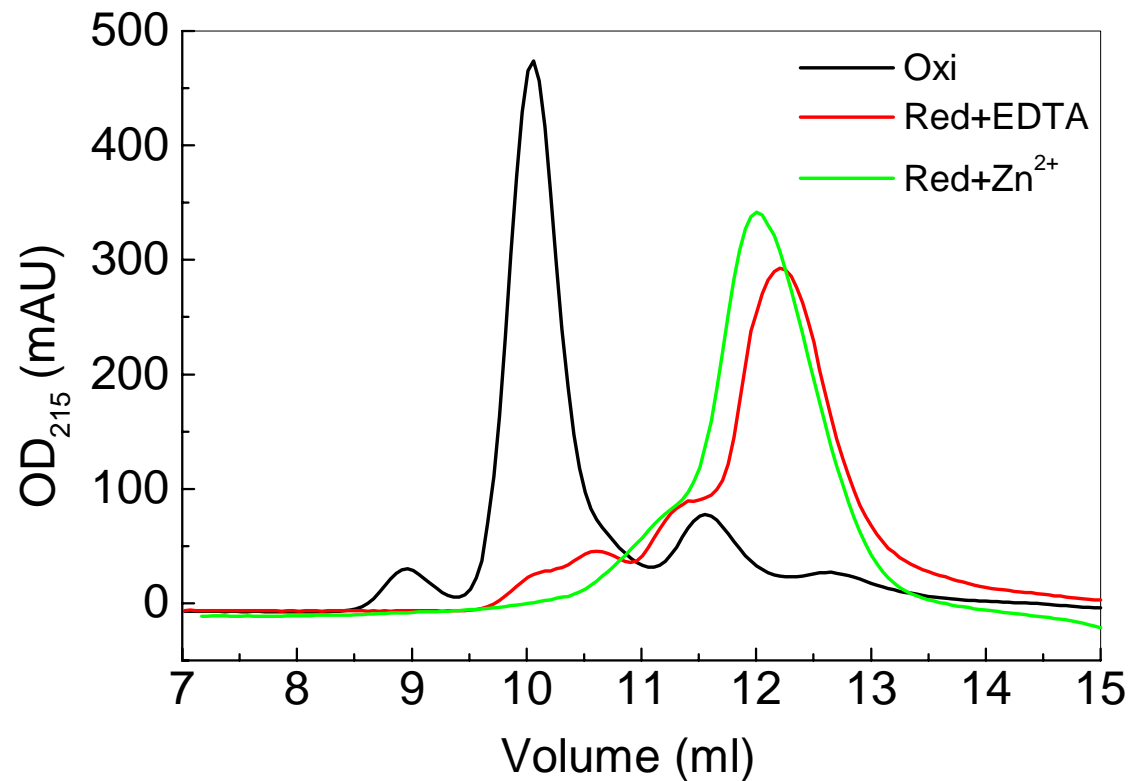


DTNB assay	GuHCl-DTT	GuHCl+DTT
Tim9	0	4.2 ± 0.5
Tim10	0	3.6 ± 0.3

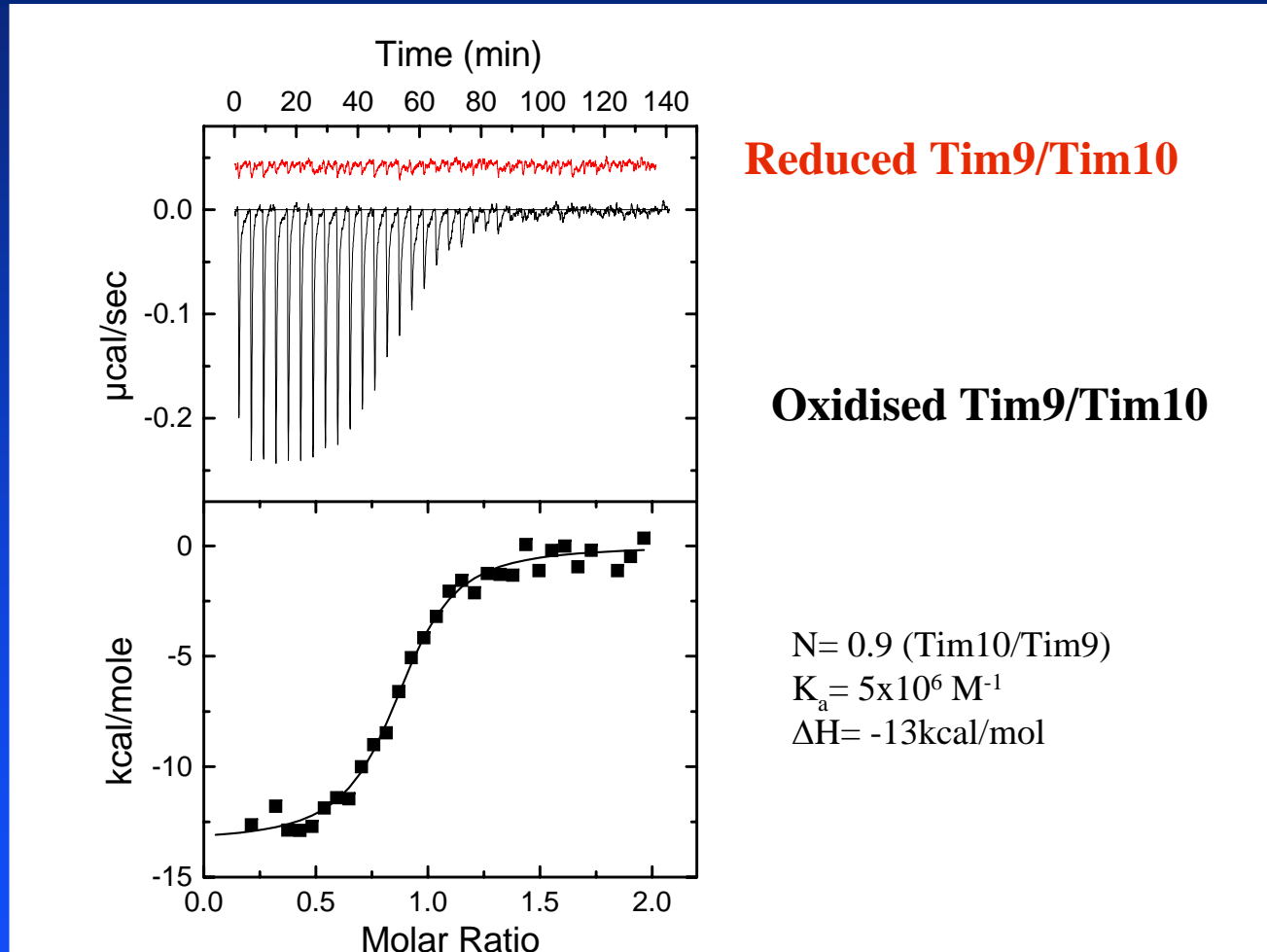
Zinc binds to reduced Tim9 and reduced Tim10, not the oxidised states



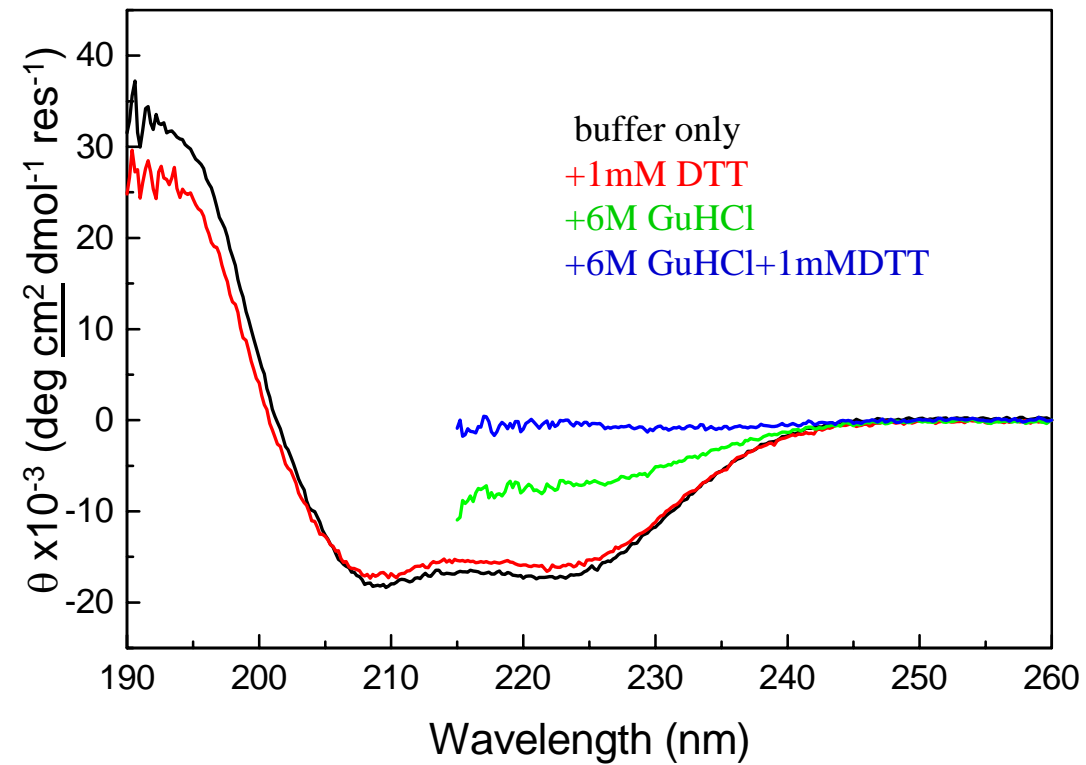
Oxidised Tim9 and oxidised Tim10 form the complex



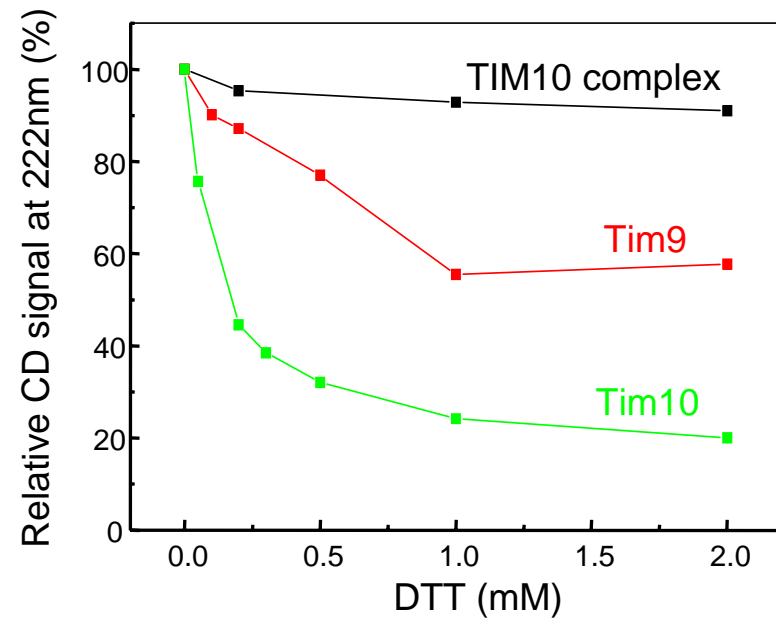
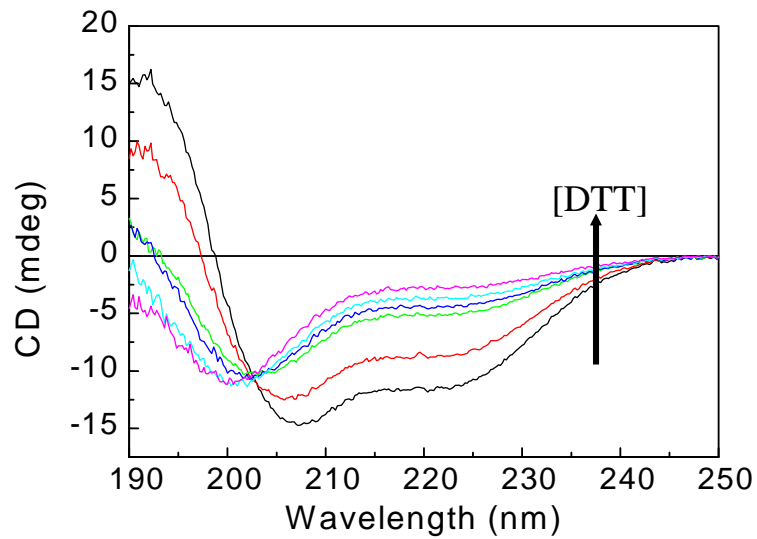
ITC study of the interaction between Tim9 and Tim10



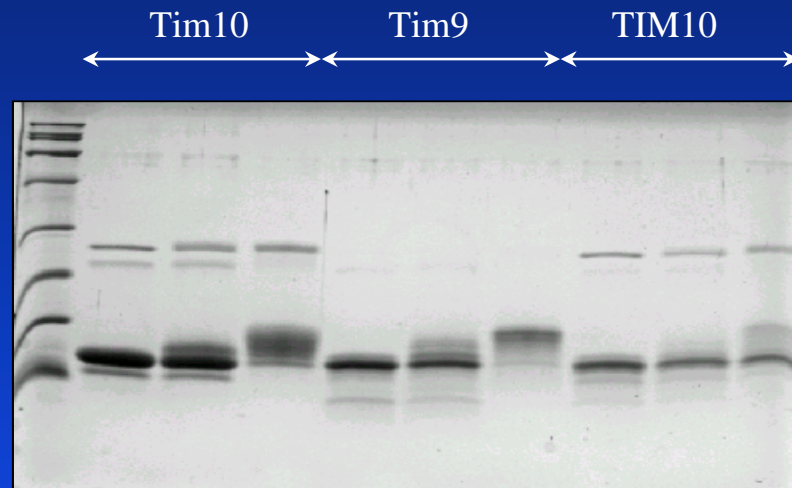
CD analysis of the TIM10 complex



DTT concentration dependence



AMS and DTNB assay for TIM10 complex



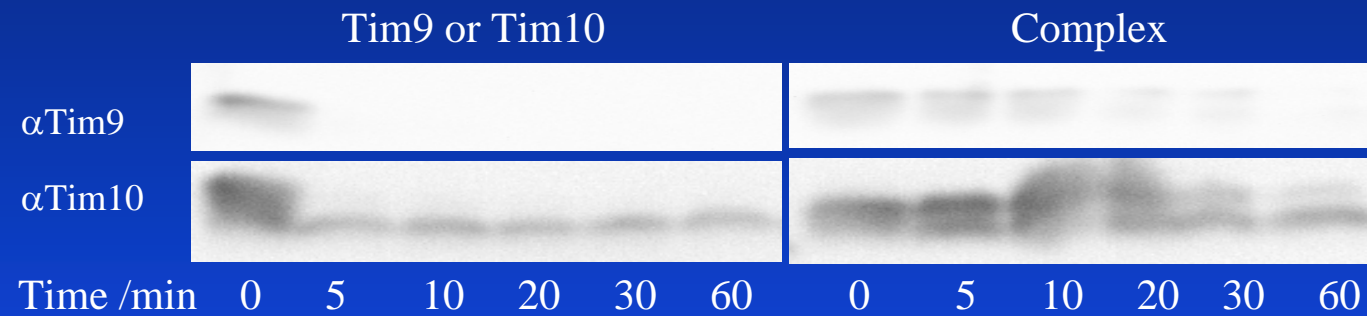
TCEP	-	-	+	-	-	+	-	-	+
AMS	-	+	+	-	+	+	-	+	+

DTNB assay of the complex:

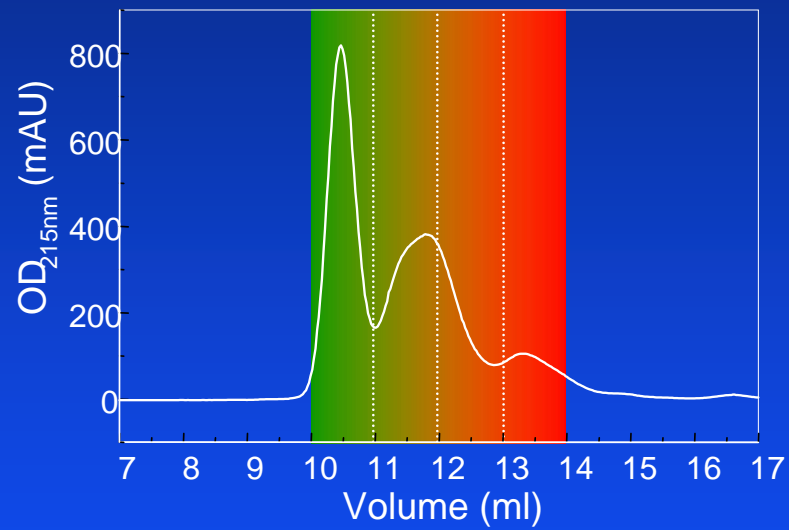
GuHCl-DTT: 0

GuHCl+DTT: 8.2 ± 0.5 per Tim9/10

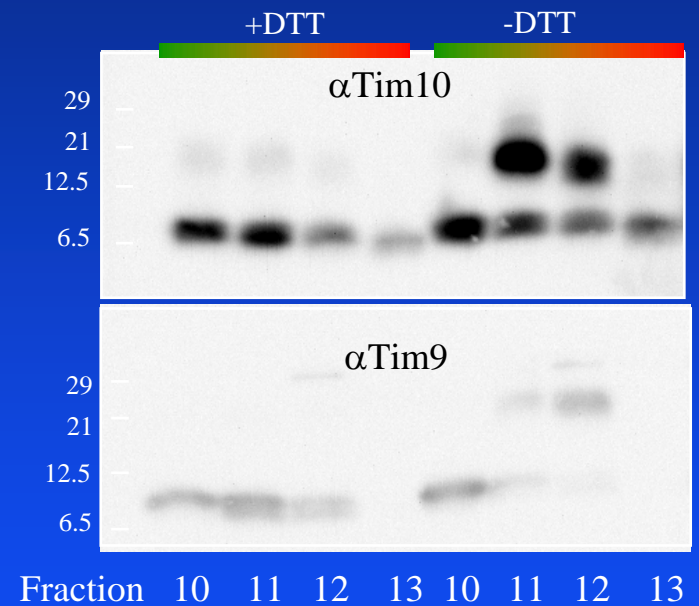
The TIM10 complex is more stable against trypsin digestion than the individual proteins



Inter- vs. intra- molecular disulfides: Misfolding & complex formation

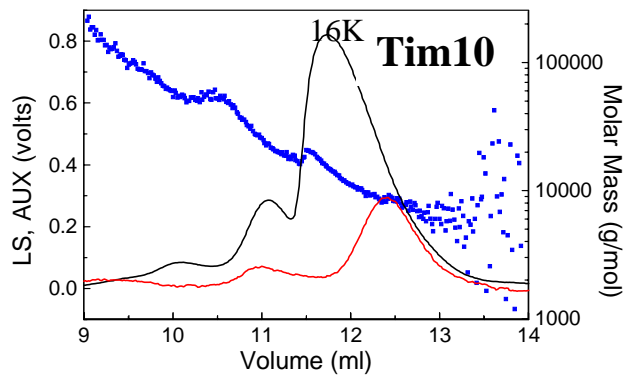
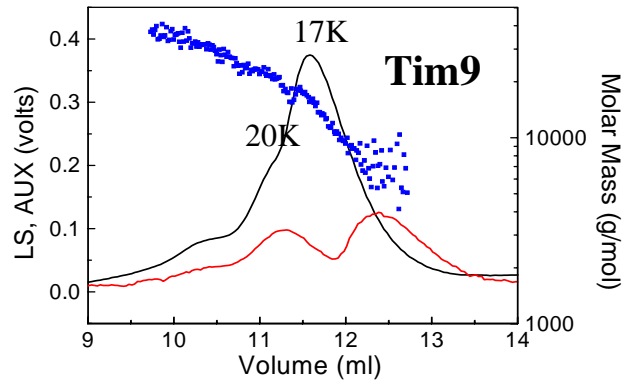


Western Blotting

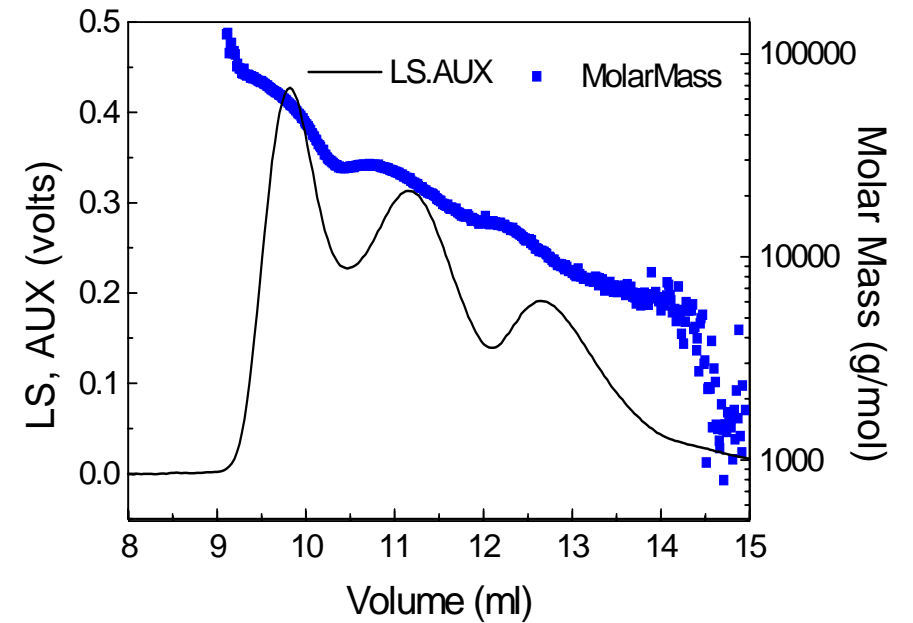


Dynamics and size analysis by multi-angle Light scattering

Individual proteins



TIM10 complex





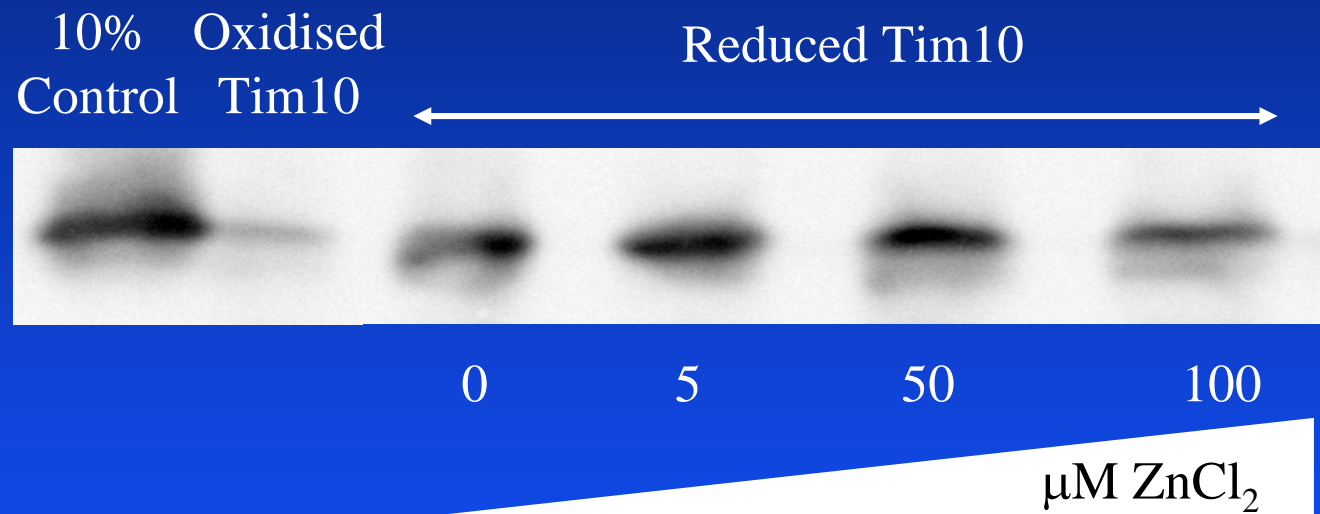
intra-molecular S-S
non-covalent
productive

inter-molecular S-S
covalent
abortive

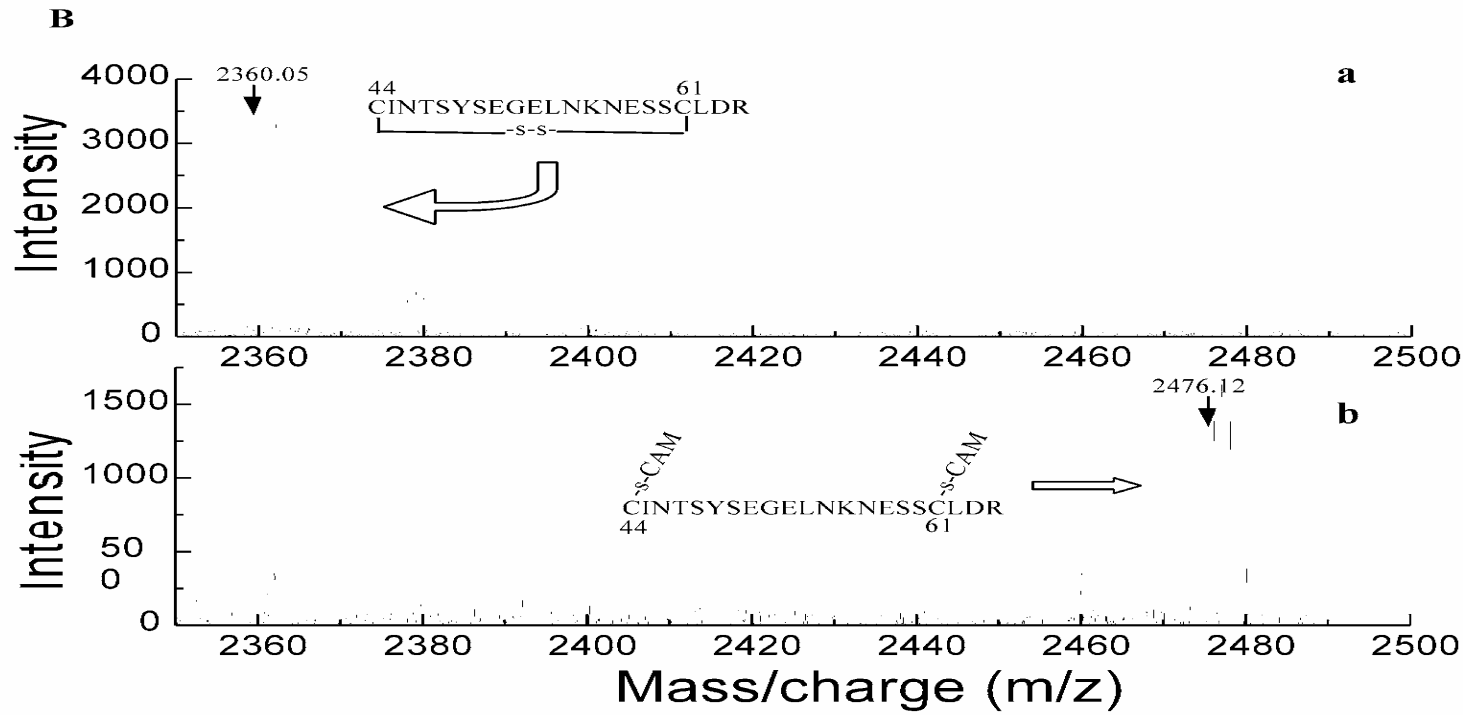
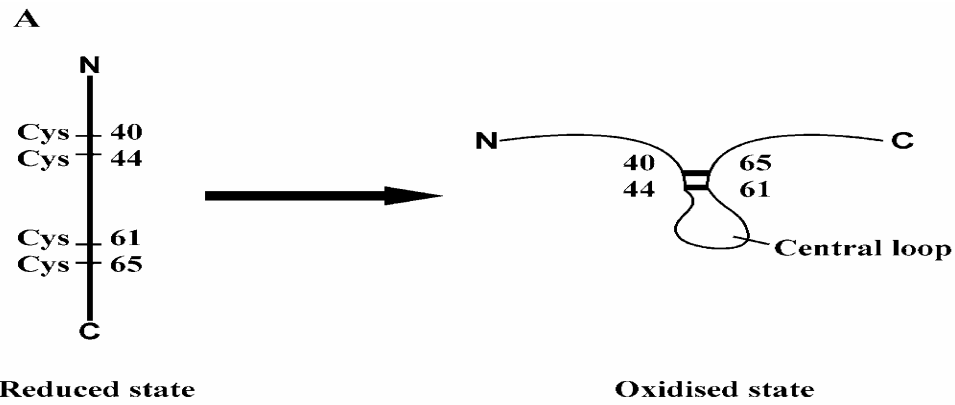


Compartment-specific redox regulation of TIM10 assembly?

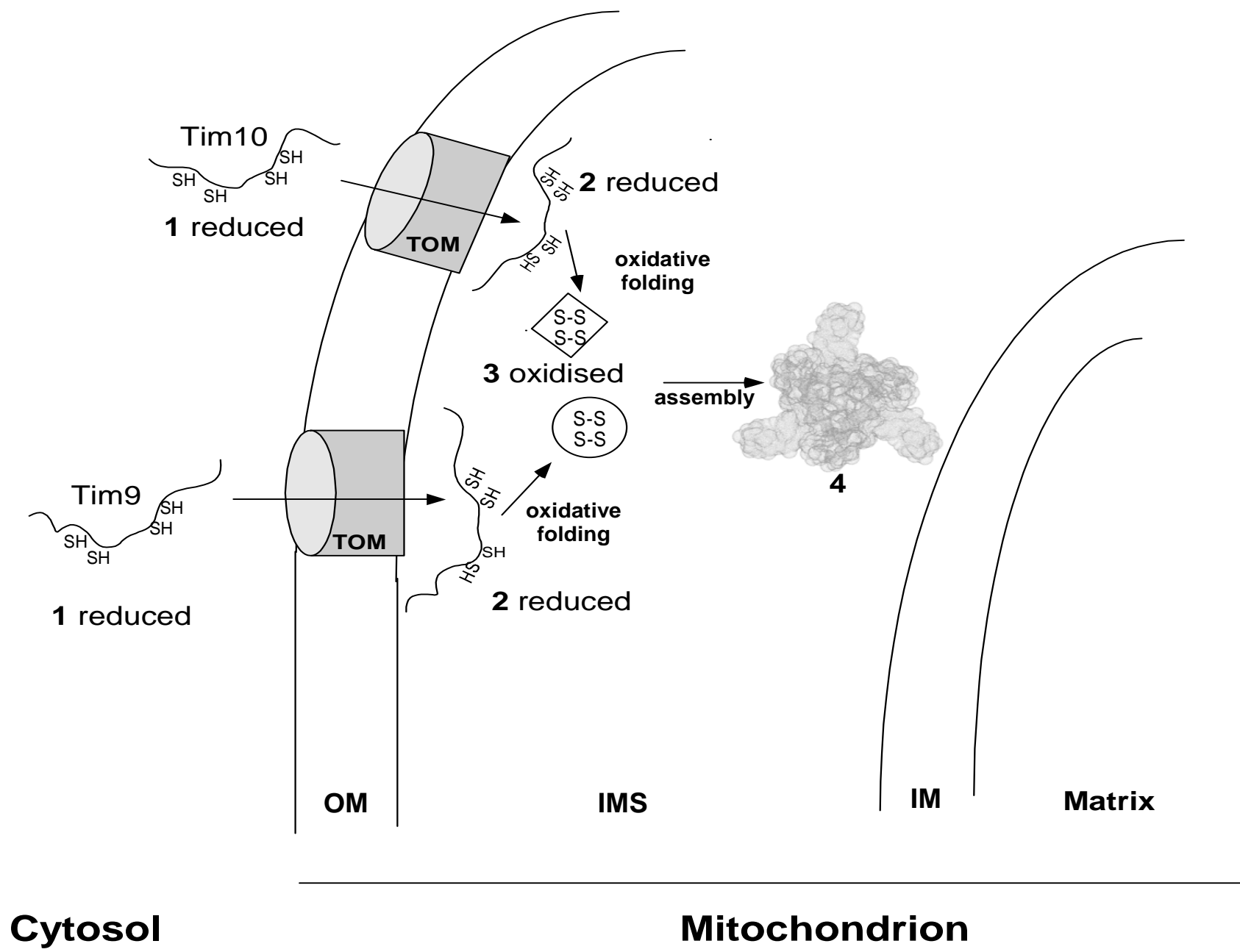
Prior Oxidation inhibits import



- Additional data:
1. NEM alkylation
 2. Cys mutants



**Oxidative folding
locks the assembly
of the TIM10 complex
in the intermembrane space**



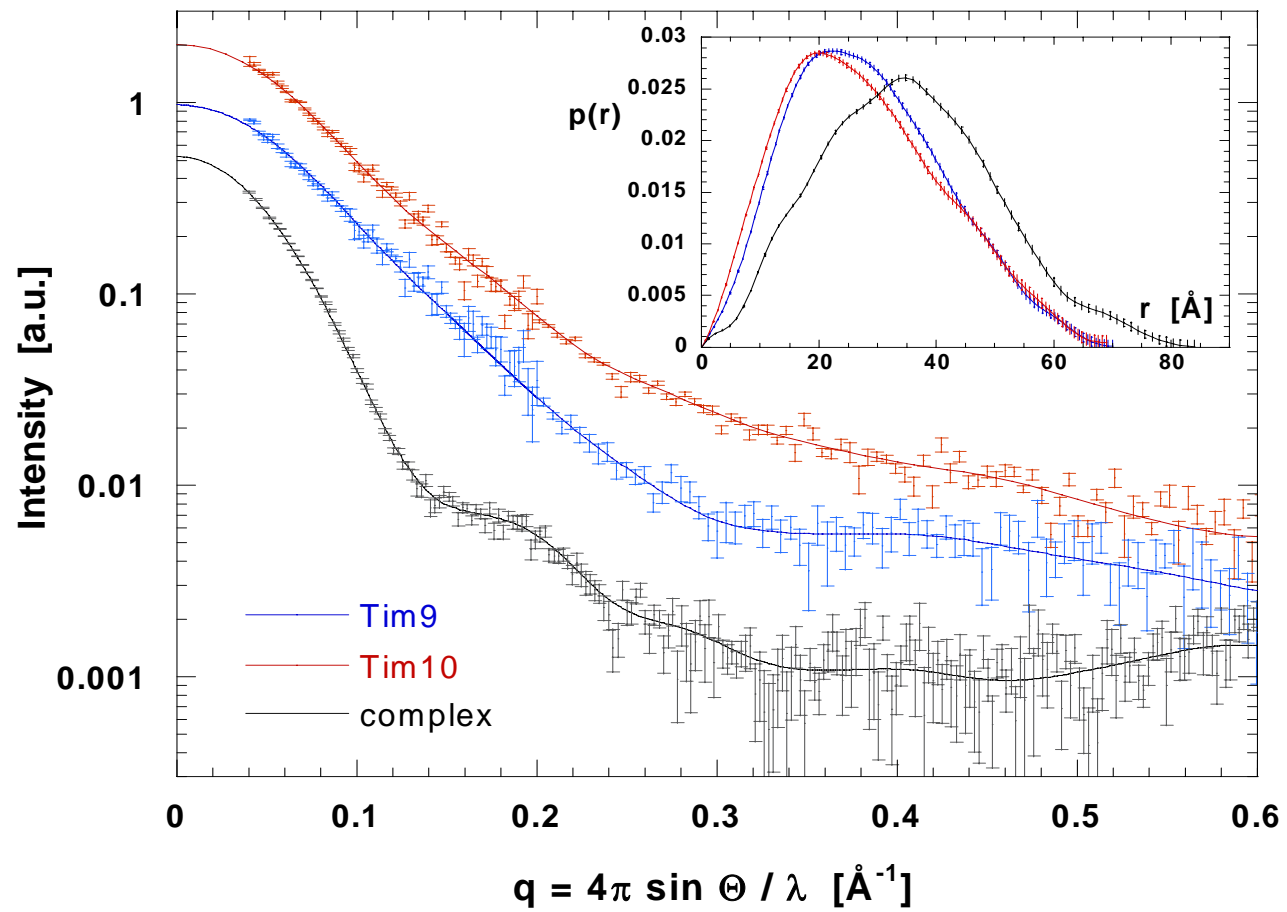
Cytosol

Mitochondrion

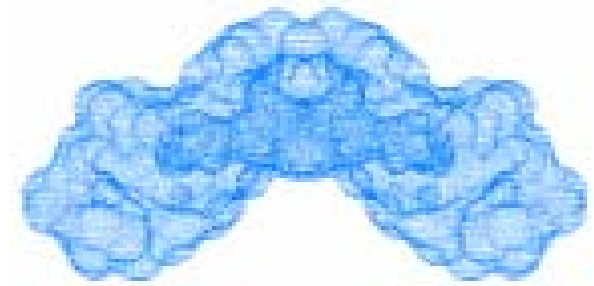
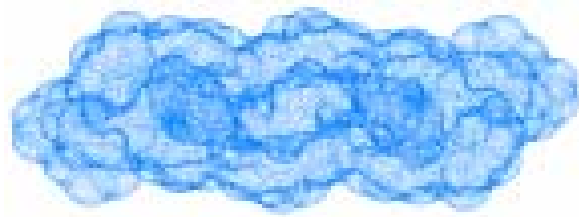
Are there distinct
functional domains
in the small Tims?

3D structure?

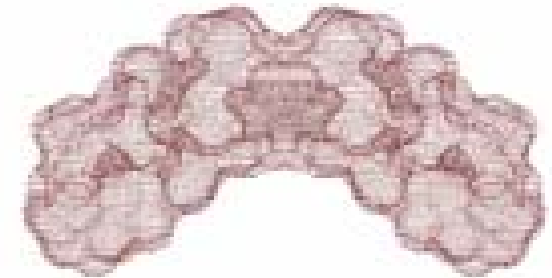
SAXS Analysis



Tim9

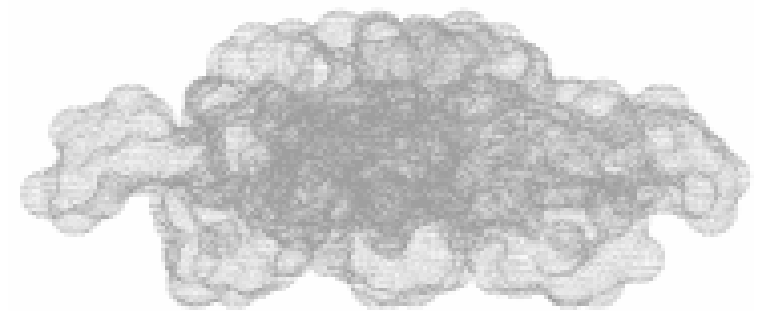
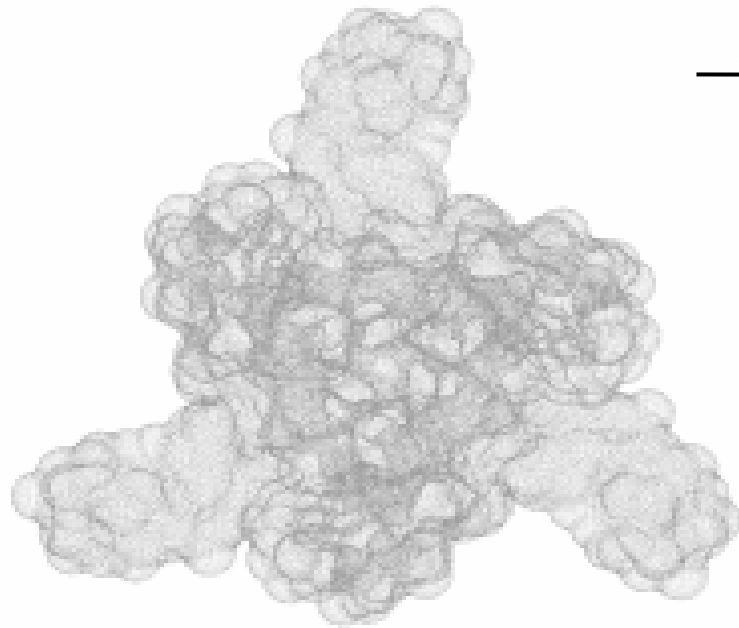


Tim10



15 Å

**Tim9-Tim10
complex**



Acknowledgments

Collaborators

- University of Manchester



Present: Stephanie Agius

Felicity Alcock

Scott Allen

Hui Lu

Maïlys Vergnolle

Past: Sabrina Dyall (UCLA, USA)

Pierre Luciano (CNRS, Marseilles France)

Peter Savory (Scotland Diagnostics)

Sarah Vial (Vertex Pharmaceuticals,
Oxford UK)

IMBB-FORTH
Catherine Baud

- Daresbury Laboratory

Günter Grossmann

- UMIST

Alexander Golovanov, Lu-Yun Lian

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